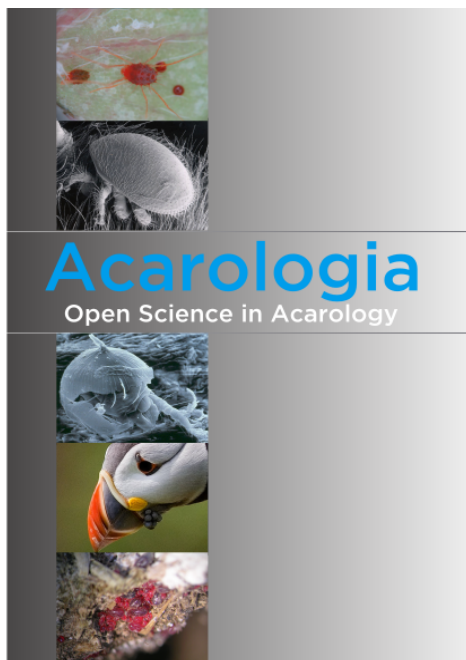




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Effect of pollen of different plant species on the oviposition of two phytoseiid mites (Acari: Phytoseiidae) commonly found in citrus orchards in the Brazilian Amazonia

Camila Tavares Ferreira^a, Cristiane Krug^b, Gilberto José de Moraes^a

^a Depto. de Entomologia e Acarologia, Escola Superior de Agricultura Luiz de Queiroz (ESALQ), Universidade de São-Paulo (USP), Piracicaba, SP, Brazil.

^b Depto. de Entomologia, Empresa Brasileira de Pesquisa Agropecuária (Embrapa Amazônia Ocidental), Manaus, AM, Brazil.

Original research

ABSTRACT

The suitability of the use of pollen to foster the control of citrus pests by facilitating the maintenance of predatory mites in the area has been extensively studied. The aim of this study was to evaluate the effect of pollen of 13 plant species commonly found in Manaus region (Brazilian Amazonia) on the oviposition of *Amblyseius aerialis* and *Iphiseiodes zuluagai*, phytoseiid species abundant in citrus orchards in that region. The study was conducted under laboratory conditions, using mostly pollen grains recently collected in the field. For comparison, a commercially available pollen (ECOPolen) and pollen obtained from two bee species were also included in the study. For about 65% of the evaluated pollen types, the average oviposition was low (less than 0.4 egg/female/day). These differed from other pollen types by the speculate wall (sunflower) or high hygroscopic property (*Agave* sp., *Amaryllis* sp., *Citrus sinensis*, *Helianthus annuus* and *Turnera ulmifolia*), or were collected by *Apis mellifera*. Highest oviposition levels of *A. aerialis* were obtained on pollen of *Typha dominguensis*, *Elaeis guineensis*, *Cocos nucifera* and the commercial pollen (1.9, 1.5, 0.9 and 0.9 eggs/female/day, respectively). For *I. zuluagai*, highest levels of oviposition were obtained on *E. guineensis*, *C. nucifera*, *T. dominguensis*, and *Elaeis oleifera* (1.6, 1.5, 1.2 and 0.9 egg/female/day, respectively). These results suggest that *C. nucifera*, *E. guineensis* and *E. oleifera*, found in the vicinity of citrus orchards in Manaus region, may play a role in maintaining predatory mite populations in such orchards. Complementary studies are needed to investigate the feasibility of maintaining those plants near orchards to increase the abundance of the predators.

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Corresponding author
Camila Tavares Ferreira:
kmlatf@hotmail.com

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Keywords predatory mites; biological control; conservation; pollen; alternative food

Introduction

Phytoseiids are important biological control agents of several pests in different crops. This large family contains more than 2,700 species (Demite *et al.* 2017), some of which have been commercially used for pest control. They are mostly found on plants, feeding mainly on phytophagous mites, but some species also feed on pollen or small insects (McMurtry *et al.* 2013). In fact, *Euseius* and *Iphiseiodes* species can be found in high abundance when the only food source available is pollen (McMurtry *et al.* 2013).

In a study conducted in Manaus region, of the Brazilian Amazonia, Bobot *et al.* (2011) reported ten species of phytophagous mites from citrus plants, belonging to the families

Eriophyidae, Tenuipalpidae and Tetranychidae. In a more recent study conducted in the same region, six species of phytophagous mites were reported by Ferreira *et al.* (2018), all of which also reported by Bobot *et al.* (2011).

Several species of predatory mites were collected in the study conducted by Ferreira *et al.* (2018), including eight phytoseiid species. The most numerous species were *Amblyseius aerialis* Muma and *Iphiseiodes zuluagai* Denmark & Muma, corroborating the results of Bobot *et al.* (2011).

Phytophagous mites are still not considered pests in citrus orchards in the Brazilian State of Amazonas, which could be due to the effectiveness of the prevailing predatory mites, although other factors may also play an important role. Pollen from native plants of Amazonia could potentially support phytoseiid populations in citrus orchards in Manaus region.

Pollen has been used as a supplement to feed predators on crops after their preventive releases. The presence of natural vegetation in the surroundings of orchards might favour the maintenance of those predators, by naturally supplying complementary food (Tixier *et al.* 2000; Duso *et al.* 2004; Maoz *et al.* 2014). Thus, more diverse agricultural systems may provide better resources to natural enemies, favouring their performance (Kennett *et al.* 1979; Tixier *et al.* 2000; Duso *et al.* 2004).

The suitability of the use of pollen to foster the control of citrus pests by facilitating the maintenance of predatory mites in the area has been extensively studied. The aim of this study was to evaluate the effect of pollen of different plant species, most of which commonly found in Amazon region, on the oviposition of *A. aerialis* and *I. zuluagai*.

Material and methods

Origin of mites and rearings

The phytoseiid species evaluated were collected from citrus orchards in Manaus a few months before initiating the experiment. Colonies were established in the laboratory, each onto a piece of resin plate (Paviflex®, 10 x 15 cm) placed onto a piece of polyethylene foam (2 cm thick) kept moist by daily additions of distilled water, inside a plastic tray (16 x 22 x 7 cm). A cover slip was placed onto cotton fibers to serve as a shelter and oviposition sites for the predator. The colonies were fed a mixture of cattail (*Typha domingensis*) pollen and immatures of *Aleuroglyphus ovatus* (Troupeau) (Acari: Acaridae). Colonies and experimental units were maintained in a rearing chamber at 25 ± 1 °C, $70 \pm 10\%$ relative humidity and 12 h of daily photophase.

Pollen sources

Pollen of 13 plant species were evaluated, including 12 species common in that region (*Agave* sp., *Amaryllis* sp., *Citrus sinensis*, *Cocos nucifera*, *Elaeis guineensis*, *Elaeis oleifera*, *Euterpe oleracea*, *Helianthus annuus*, *Montrichardia linifera*, *Oenocarpus bacaba*, *Passiflora edulis* and *Turnera ulmifolia*) and one species not found in that region, but available commercially (cattail, *T. domingensis*). Pollen was obtained from plants available in Manaus or Belém (both in the Brazilian Amazonia), except for *Amaryllis* sp., *Helianthus* sp. and *T. domingensis*, whose pollen was obtained from other parts of Brazil. For comparison, commercially available cattail pollen (of *T. domingensis*, ECOPolen, supplied by “Empresa EControle”, Brazil), and commercially available honeybee (*Apis mellifera*: Apidae) harvested pollen, as well as field collected *Melipona seminigra merrillae* (Apidae) harvested pollen were also evaluated.

To remove pollen from Arecaceae (*C. nucifera*, *E. guineensis*, *E. oleifera*, *E. oleracea* and *O. bacaba*), whose flowers were already open when collected in the field, inflorescences were collected in fabric bags, taken to air conditioned room (about 18°C) for drying overnight, and then processed according to Cunha *et al.* (2007). For most other pollen types, the flowers were still not open; hence, flowers were collected in paper bags and transported to the laboratory,

where their peduncles were maintained in a vial with water placed onto a piece of white paper, waiting for pollen grains to fall. These were then sieved to remove impurities and packed in a vial for storage in a refrigerator at -20 °C. Pollen of *C. sinensis* and *T. ulmifolia* were offered fresh to the mites, inside the anthers, due to the difficulty in separating them from other flower structures.

Experimental procedure

Each experimental unit corresponded to a plastic Petri dish (2 cm high x 3 cm in diameter) whose base was covered with two filter paper discs overlaid by a citrus leaf disc with the abaxial surface facing up. The filter paper disks were kept moist by daily addition of distilled water.

A gravid and seemingly healthy female predator was transferred with the use of a thin brush from the stock colony to an experimental unit containing pollen grains of one of types to be evaluated onto a piece of coverslip, totalling 20 females of each predator species per treatment. The amount of pollen to be supplied was determined in preliminary tests, to ensure availability of a surplus amount to the predator. Every 24 h, the old food was replaced by new food, which prevented mould incidence. The units were closed with a PVC (Magipack®) film and examined daily under a stereomicroscope to determine the number of eggs laid by the predator, in a period of 11 days. Data from the first day were not considered in the calculations to reduce the influence of previous feeding. Each female was transferred to a new unit every five days, also with the use of a thin brush.

The effect of treatments on oviposition and survivorship were compared by Kruskal-Wallis nonparametric test, as data did not have normal distribution (Shapiro Wilk's test) or stable variance (Bartlett test). Analysis was conducted using R software (R CORE TEAM 2016).

Pollen morphology

Pollen grains was examined from photographs taken under fluorescence optical (Axio Imager M2- ZEISS) and scanning electron (LEO 435-VP-ZEISS) microscopes. In the preparation of the samples for light microscopy examination, the grains were placed in a vial containing glycerin (50% glycerin and 50% water) according to Wodehouse (1933). The material was homogenized, transferring a drop with a pipette onto a slide and covering it with a coverslip. The slide was placed in a heating plate for about 30 seconds, waiting for the grains to spread on the slide, to facilitate examination. Images were taken to measure the major axis of each grain and to evaluate the purity, based on their shape. Ten pollen grains of each sample were measured and the grains were classified as very small (< 10 µm), small (10-25 µm), median (25-50 µm), large (50-100 µm), very large (100-200 µm) and giant (> 200 µm) (Halbritter *et al.* 2018). For scanning electron microscopy examination, small amounts of pollen were fixed to the upper end of a stub with the use of double-sided adhesive tape. The grains were covered with a gold layer (conductive layer) to improve contrast of the image. Photographs were then taken to examine grain size and ornamentation (Ybert *et al.* 2012; Halbritter *et al.* 2018).

Results

Oviposition and survival

For each predator species, significant differences were observed for daily oviposition rates and survivorship on the different pollen types. Oviposition rates ranged from zero to 1.6 eggs per female, with a gap between 0.4 and 0.9 egg/ female/ day for both species (except pollen of *P. edulis* for *I. zuluagai*, 0.6 egg/ female/ day). Few pollen types promoted the production of at least 0.9 egg a day (Table 1). For *A. aerialis*, the highest oviposition rates were obtained on pollen of *E. guineensis*, field collected *T. domingensis*, *C. nucifera* and the commercial pollen of *T. dominguensis* (respectively 1.5, 1.0, 0.9 and 0.9 egg/ female/day). For *I. zuluagai*, the highest

levels were obtained on pollen of *E. guineensis*, *C. nucifera*, field collected *T. domingensis* and *E. oleifera* (respectively 1.6, 1.5, 1.2 and 0.9 egg/ female/ day). For both predators, oviposition was at most 0.4 egg/ female/ day on other pollen types.

Survivorship of *A. aerialis* was at least 90% when fed pollen of *C. nucifera*, *E. guineensis*, *E. oleifera*, *E. oleracea*, field collected *T. domingensis* and commercial pollen of *T. dominguensis* (Table 1). Survivorship of *I. zuluagai* was also at least 90% when fed pollen of *C. nucifera*, *E. guineensis*, *E. oleifera*, *E. oleracea*, *H. annuus*, field collected *T. domingensis*.

Pollen morphology

Great variation in grain size and ornamentation was observed (Table 2). Sizes ranged from small to very large, but grains of palm plants were the least variable, all classified as medium sized, although they were quite variable in ornamentation. Grains collected by *A. mellifera* consisted of pollen of more than eight plant species.

Discussion

The highest level of daily oviposition of *A. aerialis* in the present study was similar or slightly higher than that obtained by Castillo and Noronha (2008), when the predator was fed pollen of *Typha angustifolia* (1.3 eggs/female). This was expected, considering the numerous reports on the suitability of pollen of plants of this genus to phytoseiid species (McMurtry and Croft 1997; Furtado and Moraes 1998; Van Rijn and Tanigoshi 1999; Ferla and Moraes 2003; Lofego and Moraes 2005; Bellini *et al.* 2010). For *I. zuluagai* the highest level of daily oviposition was also slightly higher than that obtained by Reis *et al.* (1998) and Marques *et al.* (2014) when fed with castor bean pollen (*Ricinus communis* L.) (0.7 and 0.6 egg/female respectively).

Survivorship in the course of the experiment was rather variable between treatments, but for both predators the types of pollen promoting highest oviposition rates also promoted highest survivorship rates.

Some hypothesis could be raised in relation to the performance of the predators on some pollen types. The shared features of pollen types allowing comparatively higher daily

Table 1 Mean daily oviposition ($\pm$ standard error) and survivorship (%) ($\pm$ standard error) of two phytoseiid species on different pollen types (25 ± 1 °C, $70 \pm 10\%$ RH and 12 h daily photophase).

Pollen	Family	<i>Amblyseius aerialis</i>		<i>Iphiseiodes zuluagai</i>	
		Egg/female/day ($\pm$ SE)	Survivorship (%) ($\pm$ SE)	Egg/female/day ($\pm$ SE)	Survivorship (%) ($\pm$ SE)
<i>Agave</i> sp.	Agavaceae	$<0.1 \pm 0.02$ fg	81.3 ± 5.76 cdefg	0	89.5 ± 4.33 bcde
<i>Amaryllis</i> sp.	Amaryllidaceae	$<0.1 \pm 0.02$ g	79.5 ± 5.93 fg	0.1 ± 0.02 i	71.3 ± 6.36 g
From <i>Apis melifera</i>	-	0.1 ± 0.02 efg	85.8 ± 3.75 cdefg	$<0.1 \pm 0.02$ ghi	89.5 ± 4.28 bcde
From <i>Melipona</i> sp.	-	0.1 ± 0.02 efg	87.2 ± 4.6 bcdef	0.1 ± 0.02 gh	87.7 ± 4.18 cdef
<i>Citrus sinensis</i> (orange)	Rutaceae	$<0.1 \pm 0.01$ fg	81.8 ± 4.37 fg	$<0.1 \pm 0.01$ hi	82.2 ± 5.12 efg
<i>Cocos nucifera</i> (coconut)	Arecaceae	0.9 ± 0.07 b	94 ± 3.24 abc	1.5 ± 0.06 a	99 ± 0.9 ab
<i>Elaeis guineensis</i> (African oil palm)	Arecaceae	1.5 ± 0.04 a	99.5 ± 0.45 a	1.6 ± 0.08 a	99.5 ± 0.45 ab
<i>Elaeis oleifera</i> (American oil palm)	Arecaceae	0.4 ± 0.05 c	90.4 ± 4.34 abcde	0.9 ± 0.07 c	99.5 ± 0.45 ab
<i>Euterpe oleracea</i> (açai)	Arecaceae	0.4 ± 0.03 c	93.6 ± 2.5 abcd	0.4 ± 0.04 de	93.6 ± 3.23 abcd
<i>Helianthus annuus</i> (sunflower)	Asteraceae	0.1 ± 0.03 e	81.8 ± 5.82 cdefg	0.4 ± 0.03 ef	96.8 ± 2 abc
<i>Montrichardia linifera</i> (aninga)	Araceae	0.3 ± 0.03 efg	78.6 ± 5.12 g	0	81.3 ± 5.25 fg
<i>Oenocarpus bacaba</i> (bacaba)	Arecaceae	0.1 ± 0.02 ef	83.6 ± 4.6 cdefg	0.1 ± 0.02 hi	84 ± 5.14 def
<i>Passiflora edulis</i> (passion fruit)	Passifloraceae	0.3 ± 0.02 d	85.5 ± 3.7 defg	0.6 ± 0.05 d	87.2 ± 3.9 def
<i>Turnera ulmifolia</i>	Turneraceae	0.2 ± 0.03 d	79.5 ± 5.43 efg	0.1 ± 0.02 g	80.4 ± 5 efg
<i>Typha domingensis</i> (cattail)	Typhaceae	1.0 ± 0.03 b	99 ± 0.62 ab	1.2 ± 0.09 b	100 ± 0 a
Commercial pollen of <i>T. dominguensis</i>	Typhaceae	0.9 ± 0.05 b	98.6 ± 1.36 ab	0.3 ± 0.04 f	82.2 ± 4.7 efg

Within each column, treatments whose means are followed by a same letter are not statistically different (Kruskal-Wallis nonparametric test).

Table 2 Pollen grains characteristics evaluated in this study, according to size ($\pm$ standard error) and degree of purity, according to (Ybert *et al.* 2012; Halbritter *et al.* 2018).

Scientific name	Largest dimension (μm) ($\pm$ SE)	Classification of pollen grains (size)	Ornamentation	Degree of purity (%)
<i>Agave</i> sp.	67 $\pm$ 1.09	Large	In relief	100
<i>Amaryllis</i> sp.	68 $\pm$ 1.5	Large	With depressions	100
<i>Citrus sinensis</i>	25 $\pm$ 0.6	Small	In relief	100
<i>Cocos nucifera</i>	44 $\pm$ 1.21	Medium	In relief	100
<i>Elaeis guineensis</i>	36 $\pm$ 0.56	Medium	In relief	100
<i>Elaeis oleifera</i>	32 $\pm$ 1.5	Medium	In relief	100
<i>Euterpe oleracea</i>	28 $\pm$ 0.24	Medium	Indistinct ornamentation	100
<i>Helianthus annuus</i>	31 $\pm$ 0.42	Medium	In relief	100
<i>Montrichardia linifera</i>	106 $\pm$ 2.03	Very large	Indistinct ornamentation	100
<i>Oneocarpus bacaba</i>	34 $\pm$ 1.5	Medium	Indistinct ornamentation	97
<i>Passiflora edulis</i>	66 $\pm$ 2.95	Large	In relief	100
<i>Turnera ulmifolia</i>	45 $\pm$ 0.74	Medium	In relief	100
<i>Typha domingensis</i>	25 $\pm$ 0.49	Small	With depressions	100
<i>Apis mellifera</i>	-	-	-	-
<i>Melipona</i> sp.	15 $\pm$ 0.18	Small	Indistinct ornamentation	97

(-) Not determined

oviposition rates (at least 0.9 egg/female) were: average to small sizes and irregular surfaces (either depressions or elevations) (according to the classification of Ybert *et al.* 2012). In pollen of *H. annuus*, embossment of the exine with spicules seems to have hindered exine rupturing by both predators. During the observations, it could be noticed that predators moved quickly away from those pollen grains soon after contacting them. In a study on the effect of different pollen types on the development of *Amblyseius swirskii* Athias-Henriot, the presence of very long spines on pollen of *Hibiscus syriacus* (Malvaceae) seemed to be one of the factors preventing feeding (Goleva and Zebitz 2013). In the same study, the authors classified sunflower pollen as not suitable for *A. swirskii*, but did not report whether the presence of spicules interfered in the feeding process. In the present study, oviposition rates were low on all unornamented pollen types, but presence of ornamentation was not always associated with pollen suitability.

The highest levels of oviposition could also be related to the nutritional value of each pollen type, but not much can be mentioned in relation to the nutritive quality of the pollen types used in this study. Most of the information in this regard available in the literature refers to pollen collected by bees, probably because of the relative ease in collecting large amount of those pollen types. Because of the difficulty in collecting sufficient amounts, only pollen of *E. guineensis* could be analyzed in this study. In the analysis conducted by our request by Laboratório de Bromatologia, Departamento de Zootecnia ESALQ-USP, the composition of *E. guineensis* pollen (the pollen type providing the best results for both predator species in the present study) was exactly the same reported by Figueiredo *et al.* (2018). In fact, difference in nutritional value could be one of the main reasons for the differences in oviposition rates reported for different phytoseiids on different pollen types.

The content of crude protein is considered an adequate parameter to infer pollen nutritional quality for bees; the proportion required for their development is 20 to 23% (Pernal and Currie 2000; Somerville 2005). The proportion found in *E. guineensis* pollen (38 %) is much higher than that. Massaro *et al.* (2016) reported oviposition of 0.9 and 1.1 eggs/female/day by the phytoseiids *Amblyseius tamatavensis* Blommers and *Euseius concordis* (Chant), on pollen of *E. guineensis*. However, the study of Figueiredo *et al.* (2018) offering the same type of pollen

did not confirm its suitability for the maintenance of a colony of *E. concordis*. Pollen of *C. nucifera* also favoured the oviposition of both predators in this study, and this could also be related to its high crude protein (33.5%, according to Moura 2014).

One of the common characteristics for most pollen types promoting low oviposition levels (*Agave* sp., *Amaryllis* sp., *C. sinenses*, *H. annuus*, *P. edulis* and pollen gathered by *A. mellifera*) was the water condensation on the surface of the grains during the experiment. This may have hampered mite feeding. However, the observed condensation could have happened as a consequence of the method used in this experiment in which the experimental units were closed to prevent mites from escaping. It is conceivable that under natural conditions, with pollen grains totally exposed onto plant leaves, grains could be drier in periods of no rain.

In conclusion, of the nine pollen types obtained from plants present in or around citrus orchards in Manaus region, four (*C. nucifera*, *E. guineensis*, *E. oleifera* and *P. edulis*) promoted daily oviposition of at least 0.6 egg/ female by at least one of the predator species, supporting the hypothesis that pollen from plants found in the Amazonian region can help to maintain predatory mites in citrus orchards, theoretically enhancing their effect as natural enemies of phytophagous mites.

The mite species considered in this study are commonly found in citrus orchards throughout the country (Bittencourt and da Cruz, 1988; Sato *et al.*, 1994; Reis *et al.*, 2000; Albuquerque and Moraes, 2008; Silva *et al.*, 2013; Noronha, 2013). Regarding the distribution of plants providing the best results, *E. guineensis* is mainly grown in northern Brazil and in the Bahia State (northeast), producing abundant pollen (Cunha *et al.* 2007). *Typha domingensis* and *C. nucifera* can be found in several regions of Brazil, but *T. domingensis* is not common in the north. However, by being pollen of *T. domingensis* and *E. guineensis* commercially available and readily acceptable by those predators, they could eventually be used as alternative food sources for the maintenance of predators in citrus orchards, as suggested for other predators and pollen types on the same crop (Aguilar-Fenollosa *et al.* 2011; Maoz *et al.* 2014; Jacas *et al.* 2015). Complementary field studies are needed to investigate the technical and economic feasibility of using pollen provisioning in that region, for the control of pest mites.

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References

- Aguilar-Fenollosa E., Ibáñez-Gual M.V, Pascual-Ruiz S., Hurtado M., Jacas J.A. 2011. Effect of ground-cover management on spider mites and their phytoseiid natural enemies in clementine mandarin orchards (I): Bottom-up regulation mechanisms. *Biol. Control*, 59(2): 158-170. doi: 10.1016/j.biocontrol.2011.06.013
- Albuquerque F.A. de, Moraes G.J. de 2008. Perspectivas para a criação massal de *Iphiseiodes zuluagai* Denmark & Muma (Acari: Phytoseiidae). *Neotrop. Entomol.*, 37: 328-333. doi: 10.1590/S1519-566X2008000300013
- Bellini M.R., Araujo R.V. de, Silva E.S., Moraes G.J. de, Berti Filho E. 2010. Ciclo de vida de *Proprioseiopsis cannaensis* (Muma) (Acari: Phytoseiidae) com diferentes tipos de alimentos. *Neotrop. Entomol.*, 39: 360-364. doi:10.1590/S1519-566X2010000300008
- Bittencourt M.A.L., da Cruz F.Z. 1988. Toxicidade de produtos químicos sobre ácaros predadores (Acari: Phytoseiidae) em citros. *An. Soc. Entomol. Bras.*, 17: 249-261.
- Bobot T. da E., Franklin E., Navia D., Gasnier T.R.J., Lofego A.C., Oliveira B.M. de 2011. Mites (Arachnida, Acari) on *Citrus sinensis* L. Osbeck orange trees in the state of Amazonas, Northern Brazil. *Acta Amaz.*, 41: 557-566. doi:10.1590/S0044-59672011000400013

- Castillo A.B., Noronha A.C. da S. 2008. Estudio de los aspectos fundamentales de la biología de *Amblyseius aerialis* (Muma)(Acari: Phytoseiidae) en condiciones de laboratorio. *CítriFrut*, 25(1): 45-52.
- Cunha R.N.V., Lopes R., Dantas J.C.R., Rocha R.N.C. 2007. Procedimentos para produção de sementes comerciais de dendezeiro na Embrapa Amazônia Ocidental. Manaus: Embrapa Amazônia Ocidental, 2007. 34 pp.
- Demite P.R., Moraes G.J.D., McMurtry, J.A.; Denmark H.A., Castilho R.C. 2017. Phytoseiidae Database [access September 15, 2017]. Available from: <http://www.lea.esalq.usp.br/phytoseiidae/>
- Duso C., Malagnini V., Paganelli A., Aldegheri L., Bottini M., Otto S. (2004). Pollen availability and abundance of predatory phytoseiid mites on natural and secondary hedgerows. *BioControl*, 49(4): 397-415. doi:10.1023/B:BICO.0000034601.95956.89
- Erdtman G. 1986. Pollen morphology and plant taxonomy: Angiosperms. Brill Archive. p. 553. doi:10.1126/science.117.3030.86
- Ferla N.J., Moraes G.J. de 2003. Oviposição dos ácaros predadores *Agistemus floridanus* Gonzalez, *Euseius concordis* (Chant) e *Neoseiulus anonymus* (Chant & Baker) (Acari) em resposta a diferentes tipos de alimento. *Rev. Bras. Zool.*, 20: 153-155. doi:10.1590/S0101-81752003000100019
- Ferreira C.T., Krug C., Garcia M.V.B., Moraes G.J. 2018. Leprosis mite and other mite species (Acari) associated to orange groves in Brazilian Central Amazon. *Syst. Appl. Acarol.*, 23(3): 449-462. doi:10.11158/saa.23.3.4
- Figueiredo E.S. de, Massaro M., Carmo S. do, Moraes G.J. de 2018. Rearing system for the predatory phytoseiid *Euseius concordis* (Acari: Phytoseiidae). *Exp. Appl. Acarol.*, 74(1): 13-23. doi:10.1007/s10493-018-0212-8
- Furtado I.P., Moraes G. 1998. Biology of *Euseius citrifolius*, a candidate for the biological control of *Mononychellus tanajoa* (Acari: Phytoseiidae, Tetranychidae). *Syst. Appl. Acarol.*, 3: 43. doi:10.11158/saa.3.1.6
- Goleva I., Zebitz C.P.W. 2013. Suitability of different pollen as alternative food for the predatory mite *Amblyseius swirskii* (Acari, Phytoseiidae). *Exp. Appl. Acarol.*, 61(3): 259-283. doi:10.1007/s10493-013-9700-z
- Halbritter H., Ulrich S., Grímsson F., Weber M., Zetter R., Hesse M., Bruchner R., Svojtka M., Frosch-Radivo A. 2018. Illustrated Pollen Terminology. Springer International Publishing. 483 pp. doi:10.1007/978-3-319-71365-6
- Jacas J., Aguilar-Fenollosa E., Hurtado M., Pina T. 2015. Food Web Engineering to Enhance Biological Control of *Tetranychus urticae* by Phytoseiid Mites (Tetranychidae: Phytoseiidae) in Citrus. In: *Prospect. Biol. Control Plant Feed. Mites Other Harmful Org.*: 251-269. doi:10.1007/978-3-319-15042-0_10
- Kennett C.E., Flaherty D.L., Hoffmann R.W. 1979. Effect of wind-borne pollens on the population dynamics of *Amblyseius hibisci* [Acarina: Phytoseiidae]. *Entomophaga*, 24(1): 83-98. doi:10.1007/BF02377513
- Lofego A.C., Moraes G.J. de 2005. Taxa de oviposição dos predadores *Amblyseius acalyphus* e *Amblyseius neochiapensis* (Acari: Phytoseiidae) com diferentes tipos de alimento. *Arq. Inst. Biol. (Sao. Paulo)*, 72: 379-382.
- Maoz Y., Gal S., Argov Y., Domeratzky S., Melamed E., Gan-Mor S., Coll M., Palevsky E. 2014. Efficacy of indigenous predatory mites (Acari: Phytoseiidae) against the citrus rust mite *Phyllocoptruta oleivora* (Acari: Eriophyidae): augmentation and conservation biological control in Israeli citrus orchards. *Exp. Appl. Acarol.*, 63(3): 295-312. doi:10.1007/s10493-014-9786-y
- Massaro M., Martin J.P.I., Moraes G.J. de 2016. Factitious food for mass production of predaceous phytoseiid mites (Acari: Phytoseiidae) commonly found in Brazil. *Exp. Appl. Acarol.*, 70(4): 411-420. doi:10.1007/s10493-016-0087-5
- Marques R.V., Sarmiento R.A., Ferreira V.A., Venzon M., Lemos F., Pedro-Neto M., Erasmo E.A.L., Pallini A. 2014. Alternative food sources to predatory mites (Acari) in a *Jatropha curcas* (Euphorbiaceae) crop. *Rev. Colombiana Entomol.*, 40: 74-79.
- McMurtry J.A., Croft B.A. 1997. Life-styles of Phytoseiid mites and their roles in biological control. *Annu. Rev. Entomol.*, 42: 291-321. doi:10.1146/annurev.ento.42.1.291
- McMurtry J.A., Moraes G.J. de, Sourassou N.F. 2013. Revision of the lifestyles of phytoseiid mites (Acari: Phytoseiidae) and implications for biological control strategies. *Syst. Appl. Acarol.*, 18(4): 24. doi:10.11158/saa.18.4.1
- Moura J.T. de 2014. Importância e valor nutricional do pólen de *Cocos nucifera* para abelhas africanizadas cultivadas no litoral alagoano. Universidade Federal de Alagoas, 56 pp.
- Noronha A.C. da S. 2013. Ácaros em fruteiras na Amazônia Oriental. In: Embrapa Amaz. Orient. em An. Congr., In: Simposio Brasileiro de Acarologia, 4., 2013, Bento Gonçalves.
- Pernal S.F., Currie R.W. 2000. Pollen quality of fresh and 1-year-old single pollen diets for worker honey bees (*Apis mellifera* L.). *Apidologie*, 31(3): 387-409. doi:10.1051/apido:2000130
- R Core Team. 2016. R: a language and environment for statistical computing [Access June 25, 2016]. Available from: www.r-project.org/
- Reis P.R., Chiavegato L.G., Alves E.B., Sousa E.O. 2000. Ácaros da família Phytoseiidae associados aos citros no município de Lavras, Sul de Minas Gerais. *An. da Soc. Entomológica do Bras.*, 29: 95-104. doi:10.1590/S0301-80592000000100012
- Reis P.R., Chiavegato L.G., Alves E.B. 1998. Biologia de *Iphiseiodes zuluagai* Denmark & Muma (Acari: Phytoseiidae). *An. Soc. Entomol. Brasil* 27: 185-191. doi:10.1590/S0301-80591998000200003
- Van Rijn P.C.J., Tanigoshi L.K. 1999. Pollen as food for the predatory mites *Iphiseius degenerans* and *Neoseiulus cucumeris* (Acari: Phytoseiidae): dietary range and life history. *Exp. Appl. Acarol.*, 23(10): 785-802. doi:10.1023/A:1006227704122

- Sato M.E., Raga A., Cerávolo L.C., Rossi A.C., Potenza M.R. 1994. Ácaros predadores em pomar cítrico de Presidente Prudente, Estado de São Paulo. *An. Soc. Entomol. Bras.*, 23(3): 435-441.
- Silva R.R., Silva M.J.S., Silva E.A., Serra A., Reis G.P.R. 2013. Acarofauna em pomar cítrico com ênfase em Phytoseiidae. *Magistra*, 25(3/4): 197-203.
- Somerville D. 2005. Fat Bees Skinny Bees: A Manual on Honey Bee Nutrition for Beekeepers: a Report for the Rural Industries Research and Development Corporation. Rural Industries Research and Development Corporation.
- Tixier M.-S., Kreiter S., Auger P. 2000. Colonization of vineyards by phytoseiid mites: their dispersal patterns in the plot and their fate. *Exp. Appl. Acarol.*, 24(3): 191-211. doi:10.1023/A:1006332422638
- Wodehouse R.P. 1933. Preparation of pollen for microscopic examination. *Bull. Torrey Bot. Club.*, 417-421. doi:10.2307/2480494
- Ybert J.-P., Carvalho M. de, Scheel-Ybert R., Rezende R.C. de 2012. Dicionário temático de morfologia esporopolínica. Museu Nacional. 100 pp.